Stereospecificity in the Reaction of Tetrahydro-β-carboline-3-carboxylic Acids with Isocyanates and Isothiocyanates. **Kinetic vs Thermodynamic Control**

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Benzodiazepines possessing anxiolytic, anticonvulsant, sedative-hypnotic, and myorelaxant effects are among the most widely prescribed drugs in medicine. It is now generally accepted that these compounds produce their pharmacological actions by binding to specific sites on the GABA receptor complex.¹⁻⁵ Only a small number of ligands chemically different from the benzodiazepines bind to the benzodiazepine receptor (BzR) with high affinity.⁶⁻⁹ A case in point is the group of β -carbolines, where some derivatives¹⁰⁻¹⁶ have been related to the endogenous ligand of the benzodiazepine receptor. Depending upon their intrinsic activity profiles, β -carbolines are classified as agonists, inverse agonists, antagonists, and partial agonists, making the β -carboline structure an important basis for the design of new benzodiazepine-related drugs.

In the course of a program to design new drugs acting on the central nervous system (CNS), we were interested in the synthesis of a new type of compounds 1 and 2 combining tetrahydro-*B*-carboline and hydantoin or thiohydantoin skeletons. In a previous paper¹⁷ we reported the preparation of the molecules 1 (X = 0) from isocyanates and 1-substituted-1,2,3,4-tetrahydro- β -carboline-3-carboxylic acid 3, obtained by Pictet-Spengler condensation. Nevertheless, no stereochemical control of the reaction has been studied until now.

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- Instituto de Ciencia y Tecnología de Polímeros (C.S.I.C.).
- (1) Skolnick, P.; Paul, S. ISI Atlas Sci. Pharmacol. 1988, 2, 19. (2) Gardner, C. R. Prog. Neurobiol. 1988, 31, 425.

(3) Squires, R., Ed. GABA and Benzodiazepine Receptors; CRC Press: Boca Raton, FL, 1988; Vols. I and II. (4) Prictchett, D.; Sontheimer, H.; Shives, B.; Ymer, S.; Kettenmann, H.; Schofield, P.; Seeburg, P. H. Nature 1989, 338, 582.

(5) Prictchett, D.; Luddens, H.; Seeburg, P. H. Science 1989, 245, 1389.
(6) Braestrup, C.; Nielsen, M. Arzneim. Forsch. 1980, 30, 852.
(7) Möhler, H.; Okada, T. Br. J. Psychiat. 1978, 133, 261.
(8) Tallmann, J. F.; Paul, S. M.; Skolnick, P.; Gallager, D. W. Science 20, 207.

- 1980. 207. 274.

 Muller, W. E. Pharmacology 1981, 22, 153.
 Braestrup, C.; Nielsen, M.; Olsen, C. E. Proc. Natl. Acad. Sci. U.S.A. 1980, 77, 2288.

(11) Stephens, D. N.; Shearman, G. T.; Kehr, W. Psychopharmacology (Berlin) 1984, 83, 233

(12) (a) Braestrup, C.; Honoré, T.; Nielsen, M.; Petersen, E. N.; Jensen, H. L. Biochem. Pharmacol. 1984, 33, 859. (b) Loscher, W.; Schneider, H.; Kehr, W. Eur. J. Pharmacol. 1985, 144, 261. (c) Wettstein, J. G.; Spealmann, R. D. J. Pharmacol. Exp. Ther. 1987, 240, 471. (d) Hagen,

- T. J.; Skolnick, P.; Cook, J. M. J. Med. Chem. 1987, 30, 750.
 (13) Corda, M. G.; Blaker, W. D.; Mendelson, W. B.; Guidotti, A.; Costa,
 E. Proc. Natl. Acad. Sci. U.S.A. 1983, 28, 2072.
- (14) Ninan, P. T.; Insel, T. M.; Cohen, R. M.; Cook, J. M.; Skolnick,
 P.; Paul, S. M. Science (Washington, D.C.) 1982, 218, 1332.

(15) Dorow, R.; Horowski, R.; Paschelke, G.; Amin, M.; Braestrup, C. Lancet 1983, 2, 98.

(16) File, S.; Lister, R. G.; Nutt, D. J. Neurosci. Lett. 1982, 32, 165. (17) Braña, F. M.; Garrido, M.; López-Rodríguez, M. L.; Miguel, P.; Morcillo, M. J.; Riaño, A. J. Heterocycl. Chem. 1990, 27, 703.



In keeping with our interest in the central role that the stereochemical control could play in the pharmacological profile of this type of compounds, we report here a detailed study of the stereochemistry in the reaction of 3 with isocyanates and isothiocyanates.

Initially we chose to investigate the reaction of *cis*-1substituted-1,2,3,4-tetrahydro-\beta-carboline-3-carboxylic acids 3 with isocyanates in refluxing acetone¹⁷ and acetone/ DMSO. Both conditions provided the (\pm) -cis diastereomer 1. The stereochemistry of 1 was identified by ¹³C NMR at C5 and C11a using the method of Cook et al.¹⁸ With this in mind, it could be expected that the treatment of 3 with isothiocyanates could provide the cis isomer of β -carboline-thiohydantoin rings. However when we carried out this reaction the trans diastereomer 2 was isolated as the only product.

The stereochemical assignment of 2a-f was based on a detailed study of their ¹³C NMR data and of related compounds (Tables 1 and 2). Thus, by examination of the chemical shifts for carbon atoms C5 and C11a of compounds of the types $1g-i^{17}$ and 2g-i, which do not exhibit *cis-trans* isomerism (R = H), it can be observed that the signals for these carbons in the thiohydantoin system were downfield ($\Delta C5$, 4.4; $\Delta C11a$, 3-3.5 ppm) relative to those of hydantoin (Table 2). Conversely, a cis-trans configurational change causes an upfield shift¹⁸ $(\Delta C5, 2-4; \Delta C11a, 3-4 \text{ ppm})$. Therefore, if the isolated isomer had been the cis, this compensation would not have been observed (Table 1).

The stereochemistry of 2 (trans by ¹³C NMR) was corroborated by an X-ray structure analysis (Figure 1). With respect to the selected plane defined by C5-N1-C16, the atoms C23 and H5 are located at a distance of -1.44 and -0.81 Å, respectively, both on the same side of this plane.

It was eventually demonstrated that the cis isomer 1e $[R = Ph, R' = Et; {}^{13}C NMR (CDCl_3) \delta 56.6 (C11a), 57.9$ (C5)] could be completely converted into the trans isomer 1e' [¹³C NMR (CDCl₃) δ 51.9 (C11a), 53.1 (C5)] on heating in refluxing acetonitrile in the presence of sodium carbonate. The trans isomer remained unaffected when treated under analogous conditions. Apparently, the trans diastereomer is formed under thermodynamic control. while the *cis* isomer is regulated by kinetic control.

These results have been confirmed by AM1 quantum mechanical calculations¹⁹ included in the MOPAC²⁰ program. Isomer 1e (cis) showed a heat of formation of 41.59 kcal/mol and isomer 1e' (trans) a heat of formation of 38.37 kcal/mol. The difference in energy (3.22 kcal/mol) reveals that the trans isomer is more stable than the cis and in the equilibrium the ratio of cis to trans is 0.4:99.6. The calculated heat of formation is also higher for the cis

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⁽¹⁸⁾ Ungemach, F.; Soerens, D.; Weber, R.; DiPierro, M.; Campos, O.; Mokry, P.; Cook, J. M.; Silverton, J. V. J. Am. Chem. Soc. 1980, 102, 6976. (19) Dewar, M. J. S.; Zoebisch, E. G.; Healy, E. F.; Stewart, J. J. P. J.

Am. Chem. Soc. 1985, 107, 3902.

⁽²⁰⁾ MOPAC 6.0, Quant. Chem. Prog. Exch. 1990, 455.

Scheme 1



Table 1.	¹³ C NMR Signal	s for C5 and C11	a (DMSO-d ₆) of 1	and 2a–f
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	1 a	1 b	1c	1d	le	1 f	2a	2b	2c	2dª	2e ^a	2 f°
C5	48.1	48.0	48.4	57.8	56.7	57.7	48.4	48.4	48.7	55.8	55.6	56.0
C11a	57.5	57.0	57.6	55.8	55.8	56.0	55.4	55.3	55.7	55.4	55.1	55.4

^a The signals for carbon atoms are interchangeable.

Table 2. ¹³ C NMR for C5 and C11a	(DMSO-	ds)	1
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	1 (R = H)				2(R = H)		
	g: R' = Me	h: $\mathbf{R}' = \mathbf{E}\mathbf{t}$	i: $R' = Ph$		g: R' = Me	h: $R' = Et$	i: $\mathbf{R}' = \mathbf{P}\mathbf{h}$
C5	37.6	37.5	37.8	C5	42.0	41.9	42.2
C11a	54.9	54.8	54.8	C11a	57.8	57.8	58.2



Figure 1. Crystal structure of compound 2c (R = Me, R' = Ph). Measured (X-ray) and calculated (AM1) torsion angles with crystallographic numbering system.

isomer of 2c compared to the *trans* isomer $(\Delta H_{cis} - \Delta H_{trans} = 3.35 \text{ kcal/mol})$. This indicates that in this case only the thermodynamically controlled product is obtained irrespective of the reaction conditions.

In summary, we have investigated the stereospecificity in the reaction of tetrahydro- β -carboline-3-carboxylic acid with isocyanates and isothiocyanates. This reaction provides easy access to compounds of current interest due to their potential biological applications.

Experimental Section

Solvents were purified and dried prior to use by distillation from an appropriate drying agent. Analytical thin-layer chromatography (TLC) was performed on precoated TLC plates (silica gel 60 F_{264} , layer thickness 0.2 mm). Column chromatography was performed with silica gel 60 (70–230 mesh).

Melting points were determined using a capillary melting point apparatus and are uncorrected. NMR spectra were obtained in DMSO- d_6 unless otherwise specified and chemical shifts are reported in ppm downfield relative to tetramethylsilane.

(-)-(1S,3S)-1-Methyl-1,2,3,4-tetrahydro- β -carboline-3-carboxylic acid (3a) was prepared according to the literature²¹ method.

(±)-cis-1-Phenyl-1,2,3,4-tetrahydro- β -carboline-3-carboxylic Acid (3b). Distilled benzaldehyde (4.9 mL, 48 mmol) was added to 5.02 g (24 mmol) of L-tryptophan dissolved in 40 mL of 0.1 N H₂SO₄, and the mixture was refluxed under N₂ for 6.30 h. The precipitated product was collected by filtration, washed with water (4 × 15 mL), and dried to yield 4.6 g (65%) of 3b as a white solid: mp 228-230 °C (EtOH); [α]_D =-2° (c = 1, CF₃-COOH); IR (KBr) 3460, 3280, 2940, 1640, 1570 cm⁻¹; ¹H NMR δ 2.95 (t, 11H, J = 12.9 Hz), 3.04 (dd, 11H, J = 15.3, 2.7 Hz), 3.53 (dd, 1H, J = 11.4, 4.2 Hz), 5.26 (s, 1H), 6.93-7.02 (m, 2H), 7.16 (d, 1H, J = 7.2 Hz), 7.32-7.44 (m, 6H), 10.33 (s, 1H); ¹³C NMR δ 24.1, 57.2, 57.7, 108.0, 111.5, 118.0, 118.8, 121.3, 126.3, 128.5,

^{(21) (}a) Brossi, A.; Focella, A.; Teitel, S. J. Med. Chem. 1973, 16, 418.
(b) Bobbitt, J. M.; Willis, J. P. J. Org. Chem. 1980, 45, 1978.

128.8, 129.8, 132.3, 136.8, 138.3, 171.9. Anal. Calcd for $C_{18}H_{16}N_2O_2$: C, 73.95; H, 5.52; N, 9.58. Found: C, 73.84; H, 5.46; N, 9.32.

General Preparation of Substrates 1 and 2. The corresponding isocyanate or isothiocyanate (9 mmol) was added to a suspension of 3 (9 mmol) in 35 mL of dry acetone and 15 mL of dry DMSO. The reaction mixture was refluxed to complete dissolution. The solvent was evaporated to dryness and the crude product was purified by column chromatography (silica gel, ethyl acetate/hexane 9:1).

cis-2,5-Disubstituted-5,6,11,11a-tetrahydro-1*H*-imidazo-[1',5':1,6]pyrido[3,4-b]indole-1,3(2*H*)-diones 1a-d,f. The physical and spectroscopic data are agreement with that reported previously.¹⁷

cis-2-Ethyl-5-phenyl-5,6,11,11a-tetrahydro-1*H*-imidazo-[1',5':1,6]pyrido[3,4-b]indole-1,3(2*H*)-dione (1e): mp 234-236 °C (lit.¹⁷ mp 232-234 °C); IR (KBr) 3360, 1765, 1710 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 1.15 (t, 3H, J = 7.0 Hz), 3.02 (dd, 1H, J = 15.0, 11.4 Hz), 3.32-3.60 (m, 3H), 4.32 (dd, 1H, J = 11.4, 4.4 Hz), 5.69 (s, 1H), 7.12-7.56 (m, 9H), 7.83 (s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 13.4, 22.4, 33.6, 56.6, 58.0, 106.8, 111.2, 118.4, 120.0, 122.6, 126.1, 127.8, 128.5, 128.7, 133.5, 136.7, 138.8, 154.6, 171.5.

trans-2-Ethyl-5-phenyl-5,6,11,11a-tetrahydro-1*H*-imidazo[1',5':1,6]pyrido[3,4-b]indole-1,3(2*H*)-dione (1e'). A solution of 1e (1.04 g, 3 mmol) and potassium carbonate (0.56 g, 4 mmol) in acetonitrile (17 mL) was refluxed for 2 h. The precipitate was removed by filtration and the filtrate was evaporated. The resulting oil was crystallized from ethanol to give 0.96 g (93%) of 1e': mp 245-246 °C; IR (KBr) 3360, 1770, 1710 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 1.19 (t, 3H, J = 7.0 Hz), 2.84 (dd, 1H, J =15.2, 11.1 Hz), 3.41-3.60 (m, 3H), 4.22 (dd, 1H, J = 11.0, 5.4 Hz), 6.23 (s, 1H), 7.15-7.30 (m, 8H), 7.54 (d, 1H, J = 7.8 Hz), 8.15 (s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 13.5, 23.3, 33.7, 51.9, 53.1, 107.8, 111.2, 118.4, 120.0, 122.8, 126.1, 128.1, 128.8, 129.0, 130.4, 136.6, 139.2, 154.7, 172.7. Anal. Calcd for C₂₁H₁₉N₃O₂: C, 73.04; H, 5.51; N, 12.17. Found: C, 73.33; H, 5.74; N, 12.25.

trans-2,5-Dimethyl-1-oxo-5,6,11,11a-tetrahydro-1*H*-imidazo[1',5':1,6]pyrido[3,4-*b*]indole-3(2*H*)-thione (2a): yield (85%); mp 240 °C (dec) (EtOH); IR (KBr) 3350, 1740, 1480 cm⁻¹; ¹H NMR δ 1.59 (d, 3H, J = 6.7 Hz), 2.76 (dd, 1H, J = 14.8, 11.1 Hz), 3.18 (s, 3H), 3.26 (dd, 1H, J = 14.8, 5.6 Hz), 4.73 (dd, 1H, J = 10.8, 5.6 Hz), 5.70 (q, 1H, J = 6.6 Hz), 7.01 (t, 1H, J = 7.9 Hz), 7.11 (td, 1H, J = 7.9, 1.2 Hz), 7.36 (d, 1H, J = 7.3 Hz), 7.46 (d, 1H, J = 7.6 Hz), 11.14 (s, 1H); ¹³C NMR δ 19.0, 22.5, 27.4, 48.4, 55.4, 104.0, 111.2, 118.1, 118.9, 121.5, 125.9, 134.1, 136.4, 173.3, 179.8. Anal. Calcd for C₁₅H₁₅N₈OS: C, 63.13; H, 5.30; N, 14.72; S, 11.21. Found: C, 63.23; H, 5.46; N, 14.66; S, 11.16.

trans-2-Ethyl-5-methyl-1-oxo-5,6,11,11a-tetrahydro-1*H*imidazo[1',5':1,6]pyrido[3,4-*b*]indole-3(2*H*)-thione (2b): yield (96%); mp 222 °C (dec) (EtOH); IR (KBr) 3320, 1740, 1470 cm⁻¹; ¹H NMR δ 1.17 (t, 3H, J = 7.1 Hz), 1.59 (d, 3H, J = 6.6 Hz), 2.77 (dd, 1H, J = 13.9, 11.5 Hz), 3.28 (dd, 1H, J = 14.9, 5.6 Hz), 3.80 (q, 2H, J = 6.9 Hz), 4.80 (dd, 1H, J = 11.0, 5.6 Hz), 5.70 (q, 1H, J = 6.6 Hz), 7.01 (t, 1H, J = 7.3 Hz), 7.10 (t, 1H, J = 7.3 Hz), 7.35 (d, 1H, J = 7.8 Hz), 7.48 (d, 1H, J = 7.8 Hz), 10.80 (s, 1H); ¹³C NMR δ 12.9, 19.0, 22.6, 35.7, 48.4, 55.3, 103.9, 111.2, 118.0, 118.9, 121.5, 125.9, 134.1, 136.4, 173.0, 179.2. Anal. Calcd for C₁₆H₁₇N₈OS: C, 64.18; H, 5.72; N, 14.03; S, 10.71. Found: C, 64.03; H, 5.77; N, 14.02; S, 11.00.

trans-5-Methyl-1-oxo-2-phenyl-5,6,11,11a-tetrahydro-1Himidazo[1',5':1,6]pyrido[3,4-b]indole-3(2H)-thione (2c): yield (90%); mp 240 °C (dec) (EtOH); IR (KBr) 3370, 1740, 1600, 1500, 1470 cm⁻¹; ¹H NMR δ 1.65 (d, 3H, J = 6.6 Hz), 3.04 (dd, 1H, J = 14.8, 11.0 Hz), 3.34 (dd, 1H, J = 14.8, 5.8 Hz), 4.99 (dd, 1H, J = 11.0, 5.8 Hz), 5.76 (q, 1H, J = 6.6 Hz), 7.03 (td, 1H, J= 7.5, 1.1 Hz), 7.13 (td, 1H, J = 7.5, 1.2 Hz), 7.36–7.54 (m, 7H), 10.72 (s, 1H); ¹³C NMR δ 19.0, 22.8, 48.7, 55.8, 104.1, 111.3, 118.2, 119.0, 121.7, 125.9, 128.8, 129.0, 129.2 133.5, 134.1, 136.4, 172.9, 179.5. Anal. Calcd for C₂₀H₁₇N₃OS: C, 69.13; H, 4.93; N, 12.09; S, 9.23. Found: C, 69.08; H, 5.03; N, 12.10; S, 9.12.

trans-2-Methyl-1-oxo-5-phenyl-5,6,11,11a-tetrahydro-1*H*imidazo[1',5':1,6]pyrido[3,4-*b*]indole-3(2*H*)-thione (2d): yield (76%); mp 258 °C (dec) (EtOH); IR (KBr) 3340, 1720, 1470, 1450 cm⁻¹; ¹H NMR δ 2.95 (dd, 1H, J = 15.0, 10.9 Hz), 3.14 (s, 3H), 3.46 (dd, 1H, J = 15.1, 6.1 Hz), 4.85 (dd, 1H, J = 10.7, 6.0 Hz), 6.87 (s, 1H), 7.03 (td, 1H, J = 7.8, 1.1 Hz), 7.10 (td, 1H, J = 8.0, 1.2 Hz), 7.29–7.57 (m, 7H), 10.99 (s, 1H); ¹⁸C NMR δ 22.9, 27.7, 55.4, 55.8, 105.9, 111.6, 118.5, 119.2, 122.0, 125.8, 128.6, 128.7, 128.8, 131.4, 137.0, 139.0, 173.3, 180.4. Anal. Calcd for C₂₀H₁₇N₈-OS: C, 69.14; H, 4.93; N, 12.08; S, 9.23. Found: C, 68.94; H, 4.97; N, 12.09; S, 9.50.

trans-2-Ethyl-1-oxo-5-phenyl-5,6,11,11a-tetrahydro-1*H*-imidazo[1',5':1,6]pyrido[3,4-*b*]indole-3(2*H*)-thione (2e): yield (65%); mp 206–208 °C (EtOH); IR (KBr) 3330, 1730, 1620, 1460 cm⁻¹; ¹H NMR δ 1.29 (t, 3H, J = 7.2 Hz), 3.09 (dd, 1H, J = 15.2, 10.8 Hz), 3.62 (dd, 1H, J = 15.2, 6.3 Hz), 3.93 (q, 2H, J = 7.2 Hz), 5.00 (dd, 1H, J = 10.8, 6.0 Hz), 7.05 (s, 1H), 7.18 (dd, 1H, J = 8.0, 1.5 Hz), 7.26 (td, 1H, J = 7.8, 1.5 Hz), 7.44–7.72 (m, 7H), 10.90 (s, 1H); ¹³C NMR δ 12.9, 22.9, 36.0, 55.2, 55.7, 105.8, 111.5, 118.4, 119.1, 121.9, 125.7, 128.4, 128.5, 128.7, 131.3, 136.9, 138.9, 173.0, 179.7. Anal. Calcd for C₂₁H₁₉N₃OS: C, 69.77; H, 5.31; N, 11.63; S, 8.87. Found: C, 69.50; H, 5.49; N, 11.71; S, 8.59.

trans-2,5-Diphenyl-1-oxo-5,6,11,11a-tetrahydro-1*H*-imidazo[1',5':1,6]pyrido[3,4-b]indole-3(2*H*)-thione (2*f*):yield (71%); mp 286–288 °C (EtOH); IR (KBr) 3370, 1740, 1600, 1500, 1470 cm⁻¹; ¹H NMR δ 3.21 (dd, 1H, J = 14.7, 11.1 Hz), 3.53 (dd, 1H, J = 15.0, 6.0 Hz), 5.05 (dd, 1H, J = 10.8, 6.0 Hz), 6.98 (s, 1H), 7.05 (dd, 1H, J = 7.2, 1.2 Hz), 7.13 (dd, 1H, J = 7.2, 1.2 Hz), 7.31–7.60 (m, 12H), 11.04 (s, 1H); ¹⁸C NMR δ 23.1, 55.4, 56.0, 105.8, 111.6, 118.4, 119.1, 121.9, 125.7, 128.7, 129.1, 131.3, 133.6, 136.9, 138.8, 172.7, 180.0. Anal. Calcd for C₂₅H₁₉N₃OS: C, 73.33; H, 4.68; N, 10.26; S, 7.82. Found: C, 73.58; H, 4.67; N, 10.40; S, 7.58.

2-Methyl-1-oxo-5,6,11,11a-tetrahydro-1*H*-imidazo[1',5':1,6]pyrido[3,4-b]indole-3(2*H*)-thione (2g): yield (84%); mp 227-229 °C (EtOH); IR (KBr) 3340, 1740, 1500 cm⁻¹; ¹H NMR δ 2.80 (dd, 1H, *J* = 14.8, 11.1 Hz), 3.17 (s, 3H), 3.27 (dd, 1H, *J* = 15.0, 5.7 Hz), 4.56-4.65 (m, 2H), 5.41 (d, 1H, *J* = 16.9 Hz), 7.00 (t, 1H, *J* = 7.5 Hz), 7.09 (t, 1H, *J* = 7.6 Hz), 7.36 (d, 1H, *J* = 7.8 Hz), 7.47 (d, 1H, *J* = 7.5 Hz), 11.06 (s, 1H); ¹³C NMR δ 22.3, 27.5, 42.0, 57.9, 104.6, 111.3, 117.9, 118.9, 121.4, 126.0, 129.3, 136.5, 173.2, 180.7. Anal. Calcd for C₁₄H₁₃N₃OS: C, 61.97; H, 4.83; N, 15.49; S, 11.82. Found: C, 61.67; H, 4.92; N, 15.16; S, 11.60.

2-Ethyl-1-0x0-5,6,11,11a-tetrahydro-1*H***-imidazo**[1',5':1,6]**-pyrido**[**3,4-b**]**indole-3(***2H***)-thione (2h):** yield (84%); mp 196–198 °C (EtOH/H₂O); IR (KBr) 3350, 1720, 1490, 1450 cm⁻¹; ¹H NMR δ 1.20 (t, 3H, *J* = 6.9 Hz), 2.82 (dd, 1H, *J* = 14.7, 11.1 Hz), 3.32 (dd, 1H, *J* = 15.0, 5.7 Hz), 3.83 (q, 2H, *J* = 6.9 Hz), 4.62 (d, 1H, *J* = 17.1 Hz), 4.69 (dd, 1H, *J* = 11.1, 5.7 Hz), 5.44 (d, 1H, *J* = 17.1 Hz), 7.03 (td, 1H, *J* = 7.2, 1.2 Hz), 7.12 (td, 1H, *J* = 7.2, 1.2 Hz), 7.39 (d, 1H, *J* = 7.8 Hz), 7.51 (d, 1H, *J* = 7.8 Hz), 11.03 (s, 1H); ¹³C NMR δ 13.0, 22.4, 35.8, 42.0, 57.8, 104.5, 111.3, 117.9, 118.9, 121.5, 126.0, 129.4, 136.5, 172.9, 180.1. Anal. Calcd for C₁₁₆H₁₅N₃OS: C, 63.14; H, 5.30; N, 14.72; S, 11.23. Found: C, 62.85; H, 5.50; N, 14.48; S, 11.18.

1-Oxo-2-phenyl-5,6,11,11a-tetrahydro-1*H*-imidazo[1',5':1,6]pyrido[3,4-b]indole-3(2*H*)-thione (2i): yield (91%); mp 220 °C (dec) (EtOH); IR (KBr) 3300, 1750, 1600, 1500, 1440 cm⁻¹; ¹H NMR δ 3.10 (dd, 1H, J = 14.7, 11.1 Hz), 3.47 (dd, 1H, J = 15.0, 5.7 Hz), 4.63–4.72 (m, 2H), 5.56 (d, 1H, J = 17.1 Hz), 7.16 (t, 1H, J = 7.5 Hz), 7.23 (t, 1H, J = 7.5 Hz), 7.38–7.62 (m, 7H), 11.10 (s, 1H); ¹³C NMR δ 22.5, 42.3, 58.2, 104.7, 111.3, 118.0, 119.0, 121.5, 126.1, 128.8, 128.9, 129.3, 133.6, 136.5, 172.7, 180.3. Anal. Calcd for C₁₉H₁₈N₃OS: C, 68.44; H, 4.53; N, 12.60; S, 9.62. Found: C, 68.19; H, 4.62; N, 12.31; S, 9.90.

X-ray Structural Analysis of 2c (C₂₀ H₁₇N₃OS); T = 22 °C; monoclinic, space group P_{2_1} with a = 9.144(8) Å, b = 10.282(1)Å, c = 18.765(2) Å, $\beta = 90.22$ (3)°; V = 1764(2) Å³; Z = 4; D_{caled} = 1.308 g cm⁻³; $\mu = 1.86$ cm⁻¹; F(000) = 728. The measurements were performed with an Enraf-Nonius CAD4 diffractomer with monochromatic Mo K α radiation; scan technique $\omega/2\theta$; 3289 unique data; 2164 unique data with $I \ge 2\sigma(I)$; R = 0.042; $R_w =$ 0.040. Atomic coordinates, bond lengths and angles, and thermal parameters have been deposited at the Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge, CB2 1EZ, UK.

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